

Limitation of PFAS Data and Recommendations to Obtain Appropriate Data for Environmental Decision Making

NJWEA

AAEES - Contaminants of Emerging Concern (CECs)

May 6, 2019

Nancy C. Rothman, Ph.D.

New Environmental Horizons, Inc.

34 Pheasant Run Drive, Skillman, NJ 08558

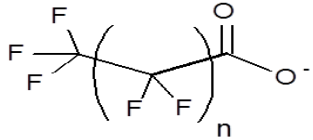
Phone: Nancy 908-874-5686

email: nrothman_neh@comcast.net

web site: www.neh-inc.com

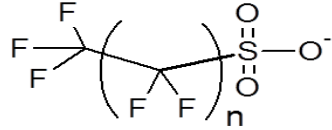
PER AND POLYFLUORINATED COMPOUNDS (PFAS/PFC)

PFCAs incl. PFOA



n=2, PFBA; n=3, PFPeA;
 n=4, PFHxA; n=5, PFHpA;
 n=6, PFOA; n=7, PFNA;
 n=8, PFDA; n=9, PFUnDA;
 n=10, PFDoDA;

PFSA incl. PFOS

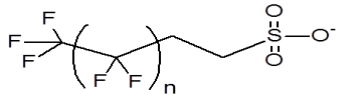


n=3, PFBS
 n=5, PFHxS
 n=7, PFOS

Poly- or perfluorinated alkyl substances (PFAS) or Perfluorocarbons (PFC) – General term for all chemicals formed from carbon chains with fluorine substituting some/all of the hydrogens on the chain

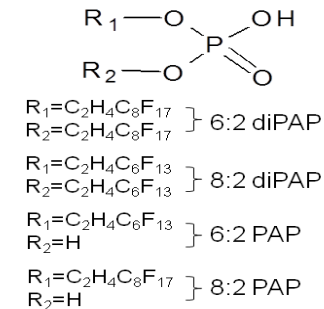
- **C-F bond** very strong
- **Unique properties** – repel water and oil, surfactant, stable
- **Diverse and complex** chemistries based on product use
- **Precursors** FTS (Fluorotelomer Sulfonate), PAP (Polyfluorinated Alkyl Phosphate Esters), PFPA (Polyfluorinated phosphonic acid), FTOH (Fluorotelomer alcohol) can all degrade to Carboxylates and Sulfonates

FTS

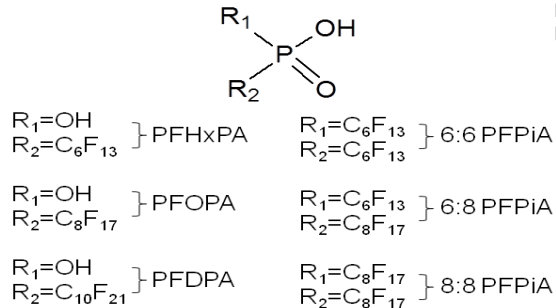


n=3, 4:2 FTS
 n=5, 6:2 FTS
 n=7, 8:2 FTS

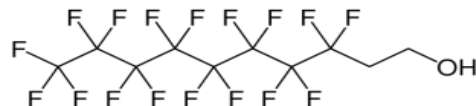
PAP, DiPAP



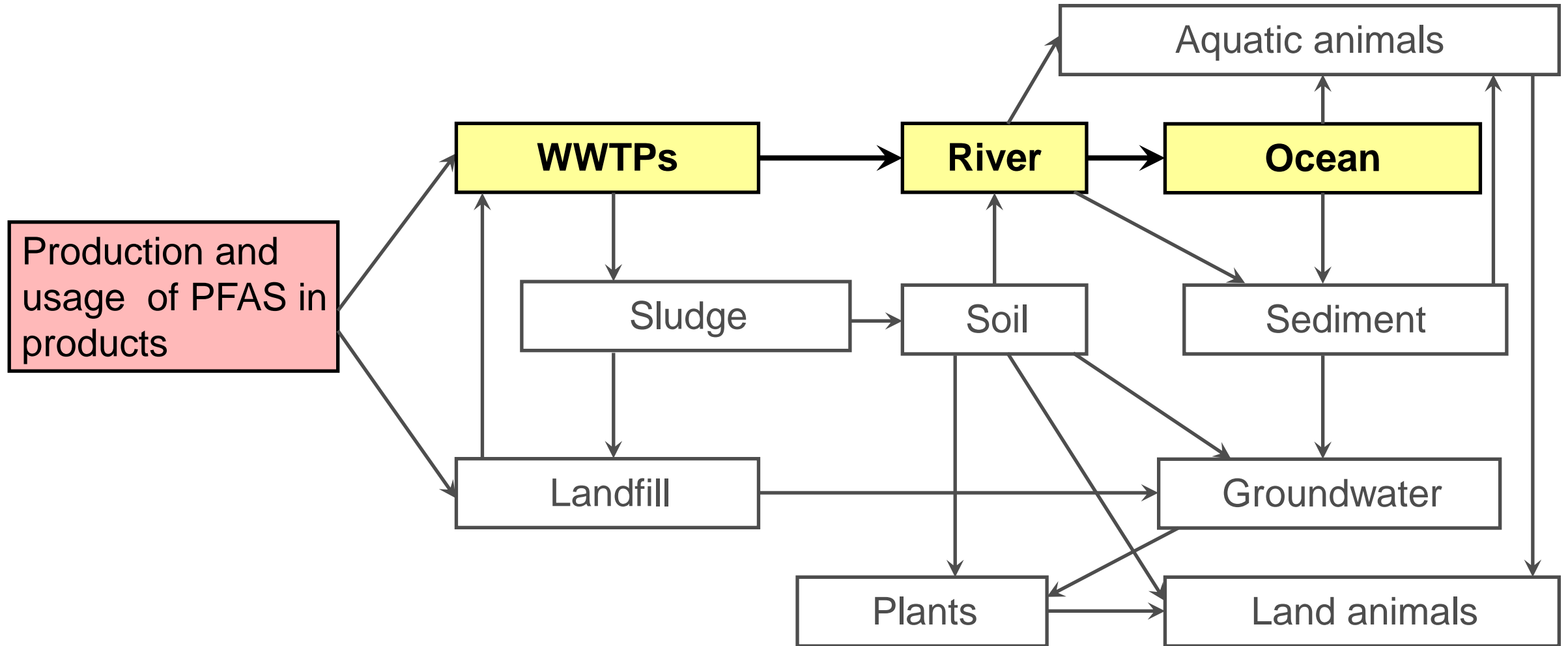
PFPA/PFPiA



FTOH



Environmental Fate of PFCs



Analysis of PFAS

USEPA Method 537.1 (version 1.0, 2018)

- Only applicable to Drinking Water samples
- No Recovery Correction
- Analyte list limited - 18 PFAS (14 PFAS required by Method 537 + 4 added compounds)
- New DW method (**Summer 2019**) - 25 PFAS includes 11 “short chain” compounds

ASTM D7979-17 & ASTM D7968 - 17a (2017)

- Non-Drinking water Aqueous & Soils
- No Recovery Correction
- 25 PFAS

Analysis of PFAS

SW-846 Method 8327 (Summer 2019)

- Direct Injection
- Non-Drinking Water Aqueous
- 24 PFAS
- No Recovery Correction

SW-846 Method 8328 (late 2019)

- Solid Phase Extraction/Isotope Dilution (SPE-ID)
- Non-Drinking Water Aqueous & Solids
- 24+ PFAS
- Recovery Correction

Lab-Specific Methods

- Modifications to the above methods
- Vary lab-to-lab

Analysis of PFAS

Total Oxidizable Precursors (TOP)

- Comparison of LCS-MS/MS results for sample pre- and post-oxidation
- Useful for evaluating Precursor potential – may be biased low

Proton Induced Gamma-ray Emission (PIGE)

- Non-destructive technique for Total Fluorine

Adsorbable Organic Fluorine /Combustible Ion Chromatography (AOF/CIC)

- Destructive technique for Total Fluorine

Analytical Issues Begin With Collection of the Samples

DoD Requirements for Personnel Collecting Samples

- No Post-it Notes; No Notebooks; No Sharpies/Markers; No Gore-Tex or Tyvek material; No Cosmetics, insect repellants, sun block, lotions worn unless 100% natural; No Waterproof material; Nitrile Gloves MUST be worn at all times

All sources of possible cross-contamination need to be eliminated and/or Evaluated

- Use Disposable Equipment if Possible
- Decontaminate equipment, if necessary, with Alconox or Liquinox using Lab Certified “PFC-free” water
- Samples should be collected in High Density Polypropylene (HDPE) with Unlined HDPE screw Caps (Teflon-lined caps MUST NOT be used)
- Trip Blanks and Field Blanks should be collected

Field Sampling Protocols to Avoid Cross-Contamination During Water Sampling for Perfluorinated Compounds (PFCs); enclosure to Navy Drinking Water Sampling Policy for Perfluorochemicals Perfluorooctane Sulfonate and Perfluorooctanoic Acid, N45 Ser/15U132432, 14 SEP 15.

Types of Data Reports

1. Summary Data Package - *Recommended*

- ◆ Narrative explaining Method of Analysis and any issues with sample receipt and analysis
- ◆ Sample Results (including FB and FD) + Surrogate recoveries
- ◆ QC results (MB, LCS, MS, & MSD or FD)
- ◆ Executed Chain-of-Custody

2. Full Deliverable – all of above + raw data

3. Result Forms/Tables only – *Not Recommended*

Specific Laboratory QA/QC For PFAS

- Sample preservation
- Sample Holding Times / Analytical Batches (≤ 20 samples)
- QC Samples required for each Analytical Batch:
 - ◆ Laboratory Reagent Blank (LRB) / Method Blank (MB)
 - ◆ Laboratory Fortified Blank (LFB) / Laboratory Control Sample (LCS)
 - ◆ Laboratory Fortified Sample Matrix (LFSM) / Matrix Spike (MS)
 - ◆ Laboratory Fortified Matrix Sample Duplicate (LFSMD) or Field Duplicate (FD)
- Surrogates added to all samples & QC prior to extraction
- Internal Standards added to all extracts prior to analysis

Recovery Surrogates vs. Isotope Dilution Surrogates

Similarities:

Added directly to the sample prior to preparation and analysis

Differences:

Recovery Surrogates

- Surrogates used to *infer* accuracy of preparation and analysis
- Internal Standards spiked prior to analysis to quantitate surrogates and target compounds

Isotope Dilution Surrogates

- Labeled Isotopes of most target compound (e.g., $^{13}\text{C}_4$ -PFOA, $^{13}\text{C}_4$ -PFOS) used for quantitation
- Loss in Isotope mirrors loss of Unlabeled compound = data are **Recovery-Corrected**

Recovery Surrogates vs. Isotope Dilution Surrogates

Non- Isotope Dilution Methods

$$\text{Compound Concentration} \equiv \frac{\text{Compound Response}}{\text{Internal Standard Response}}$$

Compound = Target PFAS

Rec. Surrogate = Recovery Surrogate

$$\text{Recovery Surrogate Concentration} \equiv \frac{\text{Rec. Surrogate Response}}{\text{Internal Standard Response}}$$

Isotope Dilution Methods

$$\text{Compound Concentration} \equiv \frac{\text{Compound Response}}{\text{ID Surrogate Response}}$$

Compound = Target PFAS

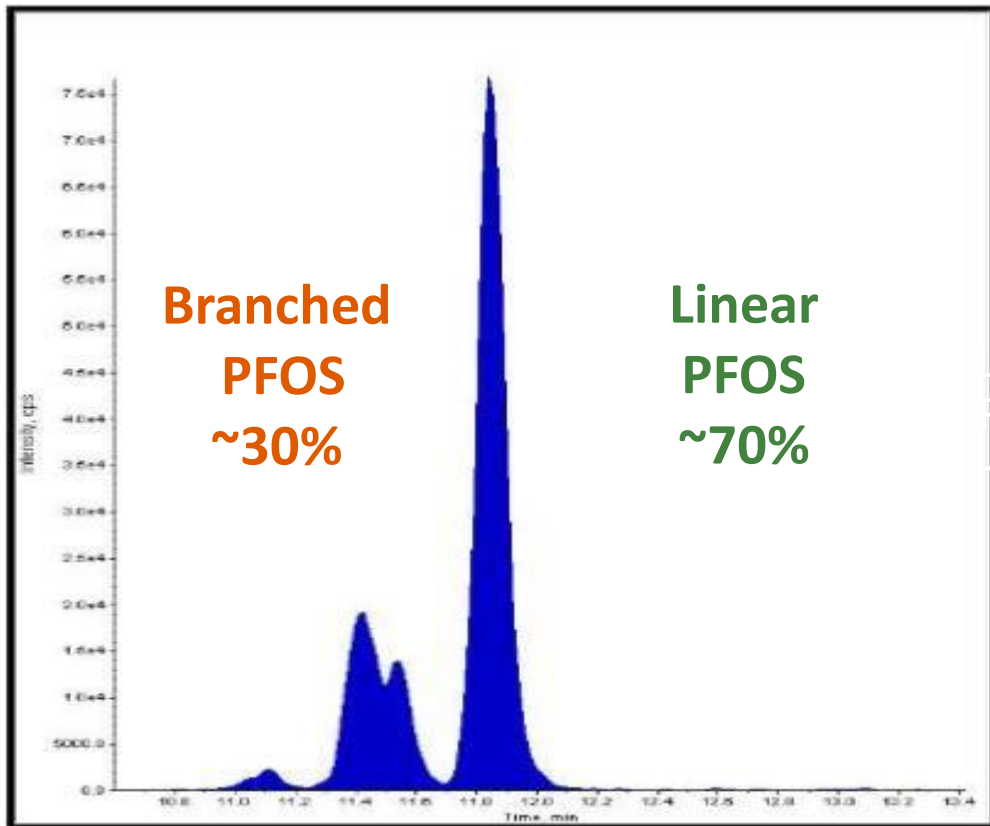
ID Surrogate = Isotope Dilution

$$\text{ID Surrogate Concentration} \equiv \frac{\text{ID Surrogate Response}}{\text{Internal Standard Response}}$$

Surrogate Recovery Problems

- Surrogate recovery below criteria: potential low bias in data
 - ◆ Due to lab error or matrix effects
- Surrogate recovery above criteria: potential high bias
 - ◆ Due to interferences or instrument issues
- **Non-Isotope Dilution Analysis** = Detected and non-detected results may be uncertain
- **Isotope Dilution Analysis** = Only compound(s) associated with Isotope affected. Uncertain whether data are biased at all since results are recovery corrected

LINEAR VS. BRANCHED ISOMERS



- Eleven *known* isomers of PFOS
- 499>80 and 499>99 transitions have different relative response factors for the linear and the branched isomers.
- Quantitative biases possible depending on standard type and MRM transitions used for quantitation
- Distribution/half lives in tissue are different between linear and branched
- Speciation is more important in research applications. Contaminant analysis issues centered around accuracy of quantitation

Riddell, N. et. al, Environ Sci. Technol. 2009 (43) 7902-7908

Data Comparability - PFAS

Factors Affecting Comparability

- Changes in Field Collection Techniques
 - Elimination or introduction of PFAS during Sampling
- Not using Isotope Dilution for Recovery Correction of data
 - Sample data may vary by $\pm 30\%$ based on Surrogate recovery acceptance limits of 70-130%
- Degradation of Precursors
 - Formation of compounds of concern over time
- Not including Branched Isomers in reporting of data
 - Historic data may not have included branched isomers
- Sensitivity differences in data sets (QLs not the same)
- Compound names being reported differently

Usability Evaluation Example

Sample	Advisory Level (ng/L)	Result (ng/L)	Surrogate %R	LCS %R	MS/MSD %R/RPD	Issue?
A	70	5 U	High	High	OK	No: Non-detect accurate as reported
B	70	66	OK	OK	%R low	Yes: result may be biased low and really >70 ng/L
C	70	63	Low	High	OK	Maybe: conflicting bias
D	70	110	Low	OK	High	No: conflicting bias but 110 >70 ng/L

Must evaluate the cumulative effect of all Quality Control to determine Usability and whether an Action Level has been exceeded

Conclusion

- Overall Quality depends on cumulative Quality from sampling through analysis
- Specifically for PFAS – Field Collection & Analytical Method differences can introduce uncertainty
- Guidelines for Evaluating Quality
 - ◆ *Data Review and Validation Guidelines for Perfluoroalkyl Substances (PFASs) Analyzed by Method 537, EPA 910-R-18-001 (November 2018)*
 - ◆ *Table B-15 of QSM 5.2 Consolidated Quality Systems Manual (QSM) for Environmental Laboratories, Version 5.2 (DOD/DOE, 2018)*
<http://www.denix.osd.mil/edqw/documents/documents/manuals/qsm-version-5-2-final-updated/>

ITRC PFAS Resource

- **Seven Fact Sheets (**available now**) and Technical Guidance Document (**late 2019**)**
- ◆ History and Use
- ◆ Nomenclature Overview and Physicochemical Properties
- ◆ Regulations, Guidance, and Advisories
- ◆ Environmental Fate and Transport
- ◆ Site Characterization Considerations, Sampling Techniques and Laboratory Analytical Methods
- ◆ Remediation Technologies and Methods
- ◆ Aqueous Film Forming Foam

<https://pfas-1.itrcweb.org/>